

## REVIEW ARTICLE

## Skin pH: From Basic Science to Basic Skin Care

Saba M. ALI and Gil YOSIPOVITCH

Department of Dermatology, Wake Forest University Baptist Medical Center, Winston-Salem, USA

The “acid mantle” is a topic not only of historical interest, but also of clinical significance and has recently been linked to vital stratum corneum function. Despite compelling basic science evidence placing skin pH as a key factor in barrier homeostasis, stratum corneum integrity, and antimicrobial defense, application of the acid mantle concept in clinical care is lacking. We review recent basic science investigations into skin pH, discuss skin disorders characterized by aberrant pH, and finally discuss practical application for preservation of the acid mantle. Recognizing factors that alter skin pH and selecting products that preserve the acid mantle is of prime importance in treating dermatologic patients. **Key words:** skin pH; acid mantle; stratum corneum; barrier homeostasis; serine protease; atopic dermatitis; acne; intertrigo; diaper dermatitis; syndets.

Accepted Nov 12, 2012; Epub ahead of print Jan 16, 2013

Acta Derm Venereol 2013; 93: 261–267.

Prof. Gil Yosipovitch, Department of Dermatology, Wake Forest University Health Sciences, Medical Center Boulevard, Winston-Salem, NC 27157, USA. E-mail: gyosipov@wfubmc.edu

Nearly a century ago, Schade and Marchionini first coined the term *Säuremantel* or “acid mantle” to describe the inherent acidic nature of the stratum corneum (SC) (1). In the last decade it has been demonstrated that skin pH largely influences barrier homeostasis, SC integrity and cohesion, and antimicrobial defense mechanisms (2–7).

In spite of mounting evidence that skin pH plays a vital role in SC function, application of the “acid mantle” concept in clinical care has lagged behind. The importance of preserving an acidic skin pH, especially in those affected by certain skin diseases, remains an under-recognized topic by practicing U.S. dermatologists. This is evident by the scarcity of low pH soaps, cleansers, and moisturizers available in the US market.

The purpose of this article is to reintroduce the subject of the “acid mantle” and provide the reader with objective evidence that skin pH is intimately linked to vital SC function. It is impossible to ignore recent compelling basic science investigations placing the role of pH in the forefront of SC function (2–6). Aberrant pH has been noted in several skin diseases and these will be reviewed. Finally, practical recommendations will be

discussed with respect to use of soaps, cleansers, and moisturizers that preserve the “acid mantle”. At the very least, we hope to provide some “pH” ood for thought.

## PHYSIOLOGIC SKIN pH

Skin pH is normally acidic, ranging in pH values of 4–6, while the body’s internal environment maintains a near-neutral pH (7–9). This creates a steep pH gradient of 2–3 units between the SC and underlying epidermis and dermis. The physiologic role of an acidic skin surface, historically was thought to be a defense mechanism against invading organisms. More recently, it has been demonstrated that several key enzymes involved in the synthesis and maintenance of a competent skin barrier are largely impacted by pH. Hence, a broader view of the importance of pH in relation to function and integrity of the skin is emerging.

## FACTORS INFLUENCING SKIN pH

A number of factors, including both endogenous and exogenous elements, affect skin pH. See Table I (10). Some of these endogenous factors will be discussed, and altered pH observed in these situations may partially explain certain clinical phenomena observed in these settings.

*Age*

Immediately after birth, skin surface pH of both full-term and preterm neonates is elevated compared to adults and older children. The mean pH value from 6 different body sites in the first day of life in full-term neonates was 7.08, which is significantly higher than in adult controls (pH 5.7) (11). pH decreases steeply in the first few days of the postnatal period and then more

Table I. Factors influencing skin pH (adopted from Yosipovitch et al. 1996 (10))

Endogenous factors	Exogenous factors
Age	Detergents, cosmetics, soaps
Anatomic site	Occlusive dressings
Genetic predisposition	Skin irritants
Ethnic differences	Topical antibacterials
Sebum	
Skin moisture	
Sweat	

gradually in the rest of the neonatal period (12–14). pH values later in infancy are similar to that of adults (15).

A decrease in pH occurs from day 3 to day 30 of the neonatal period and is most prominent in the volar forearm area compared with the forehead, cheeks, and buttocks (14). There is no disparity in pH values between different body sites in the neonate 1–2 days after birth (11). By day 90, pH is higher on the cheek and buttock and lower on the forehead and forearm (14). This apparent difference can be explained by exogenous factors, namely diaper occlusion in the buttock region and climatic factors in the exposed cheek skin (16). Eczema generally favors extensor areas in neonates, i.e. the cheeks, compared to the usual flexural distribution in adults. Extensor eczema and diaper dermatitis, commonly observed dermatoses in the infant, arise in areas with higher pH values.

A potential mechanism associated with enhanced desquamation observed in the first few days postpartum relates to the elevated pH levels. Elevated pH is known to increase activity of serine proteases, kallikrein 5 and 7, which are involved in desquamation and degradation of corneodesmosomes (5). Increased activity of these enzymes in the setting of higher pH levels likely explains the enhanced desquamation observed in the first few days postpartum, when the skin surface is more alkaline (16). Additionally, key enzymes involved in the synthesis of the permeability barrier,  $\beta$ -glucocerebrosidase and acidic sphingomyelinase, which require an acidic pH are not fully activated in the newborn period resulting in decreased skin hydration (17).

Increased skin pH and reduced buffer capacity has also been documented in skin of the elderly (9, 18, 19). Ceramide deficiency, observed in aged skin (20) has implications for barrier function and may be explained by elevated activity levels of certain enzymes that have alkaline optima. Alkaline ceramidase, which has a pH optimum of 9 and is involved in barrier lipid degradation, has higher activity in aged human skin (7).

#### *Skin site*

There are “physiologic gaps” in the acid barrier depending on skin site, particularly the interdigital spaces and intertriginous areas-axillae, groin, inframammary zone. The pH is higher in these regions compared to other skin sites (21). Higher pH in the axilla leads to colonization by certain odor-producing resident bacteria such as propionibacteria and staphylococci (22). Deodorants containing citrates reduce pH and inhibit bacterial activity (23). Candidal intertrigo also preferentially develops in the alkaline environment of the intertriginous areas.

#### *Pigmented skin*

Gunathilake et al. (4) demonstrated significantly more acidic surface pH in darkly pigmented individuals (Fitzpatrick IV–V) compared to lightly pigmented

subjects (Fitzpatrick I–II) (pH  $4.6 \pm 0.03$  vs.  $5.0 \pm 0.04$ ). Additionally, superior SC integrity and barrier function were observed in darker skin. These qualities were attributed to increased epidermal lipid content, increased lamellar body density, and lower pH in the darkly pigmented group. Serine protease activity was reduced in the more acidic environment of the darker skinned group and increased in the higher pH setting of the lightly pigmented group. Furthermore, acidification of type I–II skin with topical polyhydroxyl acids to pH levels seen in type IV–V skin enhanced barrier function in the former group to levels comparable to the darkly pigmented group (4).

### SKIN pH AND BARRIER FUNCTION

The stratum corneum’s role as a permeability barrier hinges on its hydrophobic character, lipid distribution, and organization of lipids into a series of lamellar bilayers (24). The formation of the SC barrier, specifically generation of its lipophilic components, involves several pH-dependent enzymes. Two key lipid-processing enzymes,  $\beta$ -glucocerebrosidase and acidic sphingomyelinase have pH optima of 5.6 and 4.5, respectively (7). Both are involved in the synthesis of ceramides, critical components of the permeability barrier. Activity of  $\beta$ -glucocererbrosidase is 10 times lower *in situ* at pH 7.4 than at pH 5.5 (25). Processing of lipids secreted by lamellar bodies and formation of lamellar structures require an acidic environment (26). Additionally, free fatty acids in the extracellular space form lamellar liquid crystals at pH values of 4.5–6 through partial ionization (26–28).

Investigations in both mice and human models corroborate the assertion that pH impacts barrier function. *In vivo* studies in hairless mice exposed to acetone insult or adhesive film-stripping demonstrated faster barrier function recovery in the presence of acidic buffer solution compared to neutral buffer solution (25). Similarly, blockade or knockout of secretory phospholipase A2 or the sodium-proton exchanger, both of which are involved in acidification of the SC, resulted in compromised permeability barrier homeostasis and SC integrity (2, 3). Finally, studies have shown that elevations of pH in normal skin creates a disturbed barrier, linked to increased activity of serine proteases and reduced activities of ceramide-generating enzymes (5, 6).

Recently, Hatano et al. (29) demonstrated that maintenance of an acidic SC via application of polyhydroxyl acids prevented development of hapten-induced atopic dermatitis (AD) in at-risk mice. Lowering pH in these hapten exposed mice also reduced the inflammatory TH2 response, prevented epidermal hyperplasia, reduced tissue eosinophilia, and normalized epidermal structure (29). Their findings provide intriguing implications about the use of acidic topical preparations in altering

the course of inflammatory dermatoses. Applications of polyhydroxyl acids have been shown in earlier studies to improve barrier function in both neonatal and aged rodent skin (30, 31) and even to super-normalize barrier function in normal mice (32), and in humans (4).

#### SKIN pH AND STRATUM CORNEUM INTEGRITY

pH not only influences barrier homeostasis, but also affects SC integrity, cohesion, and desquamation. Serine proteases, kallikrein 5 (SC tryptic enzyme) and kallikrein 7 (SC chymotryptic enzyme), have neutral pH optima and are intimately linked to desquamation by degrading desmoglein 1 (33–35). As pH increases, these serine proteases are activated, while the enzymes responsible for generating ceramides which have an acidic optima are inactivated compromising SC structure and function. As serine protease activity is sustained, lamellar body secretion is blocked (6, 36). See Fig. 1 for summary.

#### SKIN pH AND ANTIMICROBIAL PROPERTIES

The microflora of the skin consists of transient, temporary-resident, and permanent-resident species, including coagulase-negative staphylococci (37). Normal flora growth is optimal at acidic pH levels, whereas pathogenic bacteria, such as *S. aureus*, thrive at a neutral pH levels (38). Dermicidin, an antimicrobial peptide found in sweat, demonstrates antimicrobial activity against a variety of pathogenic microorganisms. Incubation of *S. aureus* with a sweat fraction containing dermicidin induced >90% bacteriocidal effect when buffered at pH 5.5, and only 60% when buffered at pH 6.5 (39). Chikakane & Takashashi (40) have also postulated reduced antibacterial activities of cationic substances, such as certain basic proteins, due to reduced acidity. Nitrate secreted in sweat is converted to nitrite by bacteria. Nitrite then forms reactive nitrogen species which serve as a non-specific antibacterial defense mechanism. This occurs in an acidic milieu (41).

#### SKIN pH IN DISEASE

Permeability barrier homeostasis when functioning properly imparts the skin with the capability of withstanding external insults and retaining hydration. Stratum corneum pH and permeability homeostasis are co-dependent as earlier described. Several dermatoses

characterized by disruption of the permeability barrier have altered pH and these will be discussed.

#### Atopic dermatitis

In a study of 100 children with AD, pH was observed to be significantly higher in eczematous skin and uninvolved skin in comparison to the skin of 21 healthy children (42). Others have documented similar findings of sequentially rising pH values in unaffected skin compared to perilesional skin and lesional skin in atopic patients (43, 44). Additionally, higher pH values have been measured in areas corresponding to more intense itching (44) and skin dryness in atopics (43).

Why is pH altered in atopic skin? Several contributing factors have been proposed. Free amino acids and urocanic acid, which are believed to be involved in creating the acidic milieu of the SC, are markedly reduced in atopic skin (45, 46). Filaggrin, a protein known to be deficient in AD, serves as an important precursor of free amino acids and urocanic acid. Sweat secretions, rich in lactic acid, also thought to contribute to the acid mantle, are reduced in AD (47). Finally, faulty secretion of lamellar bodies seen in AD (48), may have implications on acid pH, as exocytosis of lamellar bodies is a source of protons for SC acidification (49).

Impaired barrier function in AD can be explained in part by disturbed synthesis, excretion, and maturation of SC lipids (48, 49) processes that depend on enzymes with acidic pH optima. Aberrant lipid organization, namely increased gel phase relative to the crystalline phase of lamellar structures, has been described *ex vivo* in patients with AD (50). Lamellar liquid crystal formation occurs at pH values of 4.5–6. Serine proteases, specifically SC chymotryptic enzyme which has a pH optimum of 8 may also play a role in the pathogenesis of AD. Transgenic mice with increased serine protease activity exhibit an AD-like presentation (51). SC chymotryptic enzyme expression is dramatically increased in chronic eczema lesions (51). (Fig. 2). Moreover, serine proteases induce itch by activating PAR-2 receptors in keratinocytes and nerves in atopic skin further damaging the skin by inducing an itch scratch cycle (52).

In addition to impaired barrier, *S. aureus* colonization is a common feature of patients with AD and is considered a major pathogenetic factor in AD. Growth of staphylococcal strains is maximal at neutral pH (53) and markedly inhibited at pH values around 5 (53, 54). The 3 dimensional structure of Staphylococci enterotoxins is affected by pH. Staphylococci enterotoxin C2 has been shown to largely deviate from its normal 3-D

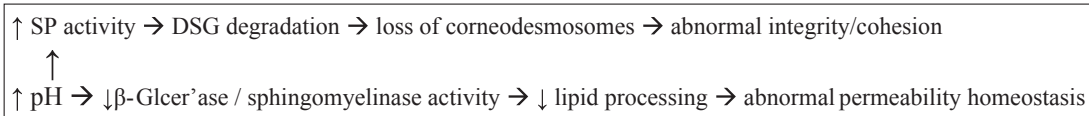


Fig. 1. Mechanism of pH altering permeability and stratum corneum integrity. SP: serine protease DSG: desmoglein.



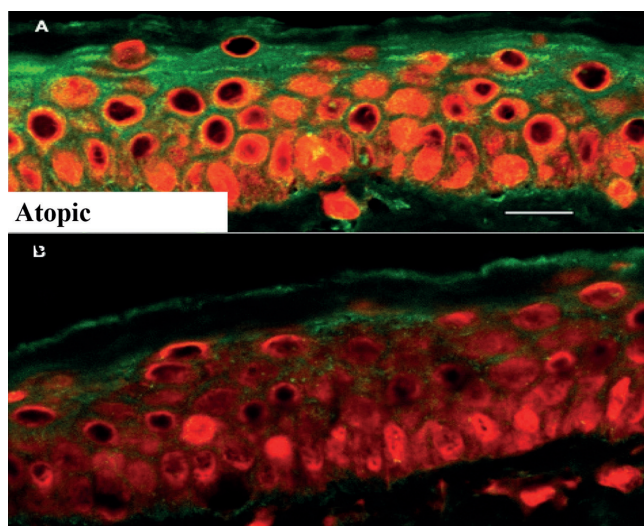


Fig. 2. Increased serine protease activity in atopic dermatitis (top) compared to normal (bottom). Orange fluorescence correlates to serine protease activity. Contributed by Dr. Peter Elias (unpublished data).

structure at pH 5 compared to pH of 8 (55). *In vitro*, the adhesion of *S. aureus* to human keratinocytes increased with increasing pH (56). See Fig. 3 for summary of pH-related events in AD.

### Ichthyosis

Öhman & Vahlquist (57), found significantly higher skin pH ( $5.3 \pm 0.7$ ) in patients with ichthyosis vulgaris compared to patients with X-linked ichthyosis ( $4.6 \pm 0.4$ ) and healthy subjects ( $4.5 \pm 0.2$ ). During tape stripping, a neutral pH of about 7 was reached in patients with ichthyosis vulgaris after half the horny layer was removed. In X-linked ichthyosis, a plateau of 6.2–6.6 in pH value was never exceeded with tape stripping. Filaggrin is known to be reduced in ichthyosis vulgaris and is also believed to play a role in acidifying the SC. Conversely, in X-linked ichthyosis, the aberration involves steroid sulphatase leading to accumulation of cholesterol sulphate and flattening of the pH gradient. Enzymes involved in desquamation are pH-dependent

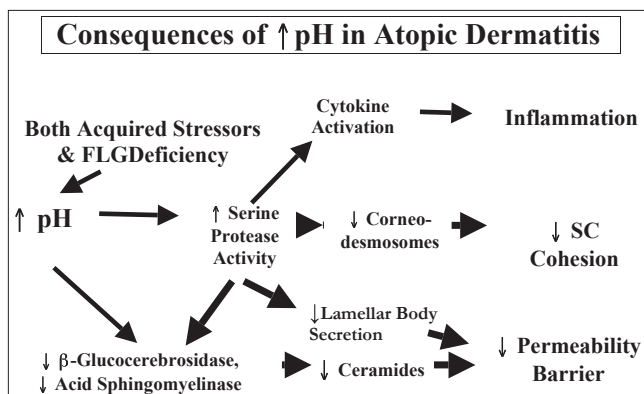


Fig. 3. Relation of skin pH in atopic dermatitis.

and alteration in pH disturbs normal desquamation (57). The use of acidic preparations with lactic acid promotes keratolysis and is effective in these ichthyoses.

### Candidal intertrigo

*Candida albicans*, a dimorphic yeast, is influenced by pH. An acidic pH favors the blastospore form and an increased pH promotes the mycelial phase (58, 59). The mycelial form is associated with pathogenicity (58, 60). In one study (61), solution with *C. albicans* was applied under occlusion to the left and right forearms of subjects, buffered at 2 different pH levels (6.0 and 4.5). Skin reactions 24 h later were more pronounced on the arm with the higher pH in 14 of the 15 subjects studied. An acidified nitrite cream has been reported to have antimycotic activity (62). Diabetics are particularly prone to develop candidal intertrigo. In a study involving non-insulin dependent diabetics, skin pH was significantly higher in the intertriginous zones of diabetics compared to intertriginous zones of healthy individuals (63). Interestingly, there was no difference in forearm pH of the two groups. Higher pH in diabetic intertriginous skin was interpreted as a possible factor promoting host susceptibility to candidal infection.

### Diaper dermatitis

A number of factors play a role in development of irritant diaper dermatitis, including prolonged exposure to urine and feces, increased hydration and occlusion, changes in skin microbial flora, and altered skin pH (64). Significant correlation between severity of diaper dermatitis and elevated skin pH in the diaper area has been demonstrated (65). Exposure of urine and feces generates ammonia and produces an alkaline environment. Alkaline pH activates fecal proteases and lipases which breakdown the skin barrier. Elevated pH also influences susceptibility to *C. albicans* as earlier described, and *C. albicans* is the microorganism most commonly associated with diaper dermatitis. Recently, Beguin et al. (66) designed an adult diaper using acidic cellulose material in order to maintain a pH of 4.5–5.5 in the diaper area. Resolution of pre-existing irritant skin lesions was noted in 8 out of 12 patients after switching to the acidified diaper design (66). Development of cleansing wipes with increased pH buffering capacity to maintain physiologic skin pH has recently been under investigation (67).

Also, tampons that are purported to reduce the usual rise in vaginal pH during menstruation have been developed and are available for purchase. Vaginal pH in healthy, pre-menopausal women ranges between 3.5–4.5. The pH of blood is 7.4 and during menstruation vaginal pH increases. RepHresh Brilliant pH tampons are currently available and employ a pH-Reducing Micro Ribbon™ and contain citric acid and L-lactide (68).

### Irritant contact dermatitis

Individuals prone to irritant contact dermatitis have been shown to have higher pH values compared to healthy individuals (69, 70). pH induced decline in SC integrity and barrier homeostasis further compounds the skin's susceptibility to injury from exposure to solvents, detergents, and mechanical forces (5).

### Tinea pedis

In one study, mean skin pH values from the foot region of subjects with tinea pedis and those without tinea pedis were compared. Foot skin pH was significantly higher in patients with tinea compared to controls (40).

### Acne

*In vitro*, *P. acnes* grows well at pH values between 6 and 6.5 and growth is markedly reduced at pH values less than 6 (71). In a study of acne-prone patients, the number of facial inflammatory lesions was compared in subjects using a conventional alkaline soap versus those using an acidic syndet bar. The number of inflammatory lesions increased in the alkaline soap group and decreased in the group using the acidic syndet at statistically significant levels by the 4<sup>th</sup> week of application (72).

### Uremics

Skin surface pH has been shown to be significantly higher in patients on dialysis compared to healthy individuals (73), despite the fact that dialysis patients have chronic acidemia. Cutaneous infections, primarily fungal infections are common in patients on hemodialysis (74). The high pH may predispose this patient population to increased mycotic infections and may suggest a possible role in uremic pruritus (73).

## PRACTICAL APPLICATION

It has been suggested that altered pH observed in the various dermatoses described earlier, is a "meaningful etiological component" and not merely an "epiphenomenon" of these conditions (75). Exposure to exogenous agents such as cleansers, creams, deodorants, and topical antibacterials affect pH and can further exacerbate underlying disease in these patients. Selection of topical agents that preserve an acidic environment seems relevant in these patients.

### Cleansers

Cleansers can be classified according to the type of surfactant used. Cleansers with non-soap-based surfactants are known as "syndets" (synthetic detergent-based bars or liquids). Syndets are generally neutral or acidic

( $\leq$ pH 7) compared to soap-based cleansers which are typically alkaline in nature (pH 10) (76). Soap-based cleansers are known to have a higher potential to irritate skin than syndets (77–82). Ananthapadmanabhan and colleagues (83) have demonstrated that high-pH solutions even in the absence of surfactants increase SC swelling and lipid rigidity. The higher pH of soap bars may be a contributing factor in the higher irritation potential of soap bars compared to syndet bars. Hand washing with soap causes the pH on the palms to increase by a mean of 3 units and remains altered 90 min after washing (7). Small and sustained pH increases, like those caused by daily use of soap-based cleansers multiple times a day, adversely influence the barrier repair mechanism (3). Baranda et al. (84) measured the pH of common cleansers (Table II). They found a significant correlation between alkaline pH of cleansers and skin irritation. In addition to information on pH of common cleansers, information regarding available low pH moisturizers is not readily accessible. Shi et al. (85) recently measured the pH of several moisturizers commonly used in the US some of which were found to have quite high alkaline pH (Table SI; available from <http://www.medicaljournals.se/acta/content/?doi=10.2340/00015555-1531>).

### Acidification of the stratum corneum

Topical alpha-hydroxy acids (AHA) are common agents used in treating disorders of keratinization. AHA, such as lactic acid have been shown to increase ceramide

Table II. pH of cleansers (Adopted from Baranda et al. (84))

Brand name	pH	Composition
Aderm	6.44	Syndet
Avecyde	3.61	Syndet
Avène	6.94	Syndet
Cetaphil	7.72	Syndet
Dove white	7.53	Syndet
Dove baby	7.0	Syndet
Dove (liquid)	5.16	Syndet
Dove pink	7.23	Syndet
Johnson's baby	11.9	Soap
Johnson's baby oat	12.35	Soap
Nivea baby creamy	12.35	Syndet <sup>a</sup>
Nivea bath care	12.21	Syndet <sup>a</sup>
Nivea bath c. Almond	12.22	Syndet <sup>a</sup>
Nivea bath c. Oat	12.30	Syndet <sup>a</sup>
Zest neutral	9.85	Soap
Zest citrus sport	9.75	Soap
Zest herbal	9.97	Soap
Zest aqua	9.89	Soap
Palmolive green	10.18	Soap
Palmolive (white)	10.23	Soap
Palmolive botanicals	10.38	Soap
Palmolive botanicals/camomile	10.13	Soap
Camay classic	10.38	Soap
Camay gala	10.36	Soap
Camay soft	10.26	Soap

<sup>a</sup>plus mineral oil.

production by human keratinocytes by 300% *in vitro* (86). In one study, twice daily application of 4% l-lactic acid formulations (pH 3.7–4.0) led to significant improvement in barrier function as measured by TEWL and reduced sensitivity to sodium lauryl sulphate (SLS) after 4 weeks (86). *In vivo* the total ceramide fraction increased significantly. The ability of AHA to increase ceramide levels is beneficial in those individuals with reduced barrier function such as atopics who have reduced levels of ceramides (87, 88). Studies have shown beneficial effects of topical acidic electrolyte water (pH 2.0–2.7) on the severity of dermatitis and *S. aureus* colonization of the skin in children (89) and adults (90). Use of AHA in irritant dermatitis appears useful as evident by reduction in sensitivity to SLS.

### Guidelines for dermatologists

In managing cutaneous diseases such as acne, AD, intertrigo, and irritant contact dermatitis the clinician has an armamentarium of prescription topicals and oral agents. Use of proper soaps and over-the-counter creams that do not compromise the acidic pH of skin should become part of the treatment regimen of these patients. In recommending the ideal body wash, soap or cleanser, one that has a pH between 4.5–6.5, similar to the normal pH of the skin, should be selected. Syndets are less irritating and are preferred. Frequent use of alkaline medicated soaps containing benzoyl peroxide, sulfur, or resorcinol antibacterials (eg, triclocarban or triclosan), although excellent in eradicating Staphylococci and gram negative bacteria have a pH of 9–10 and cause skin irritation, so daily use should be discouraged. Often, patients with intertrigo or acne, believing that their dermatoses are related to poor hygiene, overuse harsh soaps exacerbating the condition and a vicious cycle of further cleansing ensues. Proper education and recommendations on appropriate topicals is crucial in these situations. In atopics, creating an acidic pH by selecting appropriate topicals is vital in restoring a pre-existing faulty barrier as earlier discussed. One may even consider measuring skin pH in clinic as a bedside measurement, which is a non-invasive, simple test. However the current available measurements using flat glass electrodes have limitations. The glass electrode needs calibration and measurement errors could be caused by surplus of water as well as dry electrode surface. Factors that can influence the results are sweat, washing procedures that can have alkalinizing effect on skin surface pH regardless of the pH of the solution (91).

### CONCLUSION

In the last decade, the role of skin pH as a factor in vital SC function has been investigated. Likely, much remains to be learned about the complex relation of skin pH and

downstream pH dependant events. We do know that many skin diseases characterized by faulty barrier function have aberrant pH values. This should prompt the clinician to focus on preserving or restoring an acidic milieu by selecting topical agents compatible with the acid mantle.

*The authors declare no conflict of interest.*

### REFERENCES

- Schade H, Marchionini A. Der Säuremantel der Haut nach Gaskettenmessngen. *Klin Wochenschr* 1928; 7: 12–14.
- Behne MJ, Meyer JW, Hanson KM, Barry NP, Murata S, Crumrine D, et al. NHE1 regulates the stratum corneum permeability barrier homeostasis. Microenvironment acidification assessed with fluorescence lifetime imaging. *J Biol Chem* 2002; 277: 47399–47406.
- Fluhr JW, Kao J, Jain M, Ahn SK, Feingold KR, Elias PM. Generation of free fatty acids from phospholipids regulates stratum corneum acidification and integrity. *J Invest Dermatol* 2001; 117: 52–58.
- Gunathilake R, Schurer NY, Shoo BA, Celli A, Hachem JP, Crumrine D, et al. pH-regulated mechanisms account for pigment-type differences in epidermal barrier function. *J Invest Dermatol* 2009; 129: 1719–1729.
- Hachem JP, Crumrine D, Fluhr J, Brown BE, Feingold KR, Elias PM. pH directly regulates epidermal permeability barrier homeostasis, and stratum corneum integrity/cohesion. *J Invest Dermatol* 2003; 121: 345–353.
- Hachem JP, Man MQ, Crumrine D, Uchida Y, Brown BE, Rogiers V, et al. Sustained serine proteases activity by prolonged increase in pH leads to degradation of lipid processing enzymes and profound alterations of barrier function and stratum corneum integrity. *J Invest Dermatol* 2005; 125: 510–520.
- Rippke F, Schreiner V, Schwanitz HJ. The acidic milieu of the horny layer: New findings on the physiology and pathophysiology of skin pH. *Am J Clin Dermatol* 2002; 3: 261–272.
- Dikstein S, Zlotogorski A. Measurement of skin pH. *Acta Derm Venereol Suppl* 1994; 185: 18–20.
- Zlotogorski A. Distribution of skin surface pH on the forehead and cheek of adults. *Arch Dermatol Res* 1987; 279: 398–401.
- Yosipovitch G, Maibach HI. Skin surface pH: a protective acid mantle. *Cosmet Toiletries* 1996; 111: 101–102.
- Yosipovitch G, Maayan-Metzger A, Merlob P, Sirota L. Skin barrier properties in different body areas in neonates. *Pediatrics* 2000; 106: 105–108.
- Fox C, Nelson D, Wareham J. The timing of skin acidification in very low birth weight infants. *J Perinatol* 1998; 18: 272–275.
- Green M, Carol B, Behrendt H. Physiologic skin pH patterns in infants of low birth weight. The onset of surface acidification. *Am J Dis child* 1968; 115: 9–16.
- Hoeger P H, Enzmann C. Skin physiology of the neonate and young infant: a prospective study of functional skin parameters during early infancy. *Pediatr Dermatol* 2002; 19: 256–262.
- Fluhr JW, Pfistere S, Gloor M. Direct comparison of skin physiology in children and adults with bioengineering methods. *Pediatr Dermatol* 2000; 17: 436–439.
- Fluhr JW, Darlenski R, Taieb A, Hachem JP, Baudouin C, Msika P, et al. Functional skin adaptation in infancy – almost complete but not fully competent. *Exp Dermatol*



- 2010; 19: 1–10.
17. Fluhr JW, Darlenski R, Lachmann N, Baudouin C, Msika P, De Belilovsky C, Hachem JP. Infant epidermal skin-physiology: Adaptation after birth. *Br J Dermatol* 2012; 166: 483–490.
  18. Laufer A, Dikstein S. Objective measurement and self-assessment of skin-care treatments. *Cosmet Toiletries* 1996; 111: 91–98.
  19. Thune P, Nilsen T, Hanstad IK, Gustavsen T, Lövig Dahl H. The water barrier function of the skin in relation to the water content of stratum corneum, pH and skin lipids. The effect of alkaline soap and syndet on dry skin in elderly, non-atopic patients. *Acta Derm Venereol* 1988; 68: 277–283.
  20. Jin K, Higaki Y, Takagi Y, Higuchi K, Yada Y, Kawashima M, Imokawa G. Analysis of beta-glucocerebrosidase and ceramidase activities in atopic and aged dry skin. *Acta Derm Venereol* 1994; 74: 337–340.
  21. Braun-Falco O, Korting HC. Der normale pH-Wert der Haut. *Hautarzt* 1986; 37: 126–129.
  22. Korting HC, Lucacs A, Braun-Falco O. Mikrobielle Flora und Geruch der menschlichen Haut. *Hautarzt* 1988; 39: 564–568.
  23. Stenzaly-Achtert S, Schölermann A, Schreiber J, Diec KH, Rippe F, Bielfeldt S. Axillary pH and influence of deodorants. *Skin Res Technol* 2000; 6: 87–91.
  24. Elias PM, Menon GK. Structural and lipid biochemical correlates of the epidermal permeability barrier. *Adv Lipid Res* 1991; 24: 1–26.
  25. Mauro T, Holleran WM, Grayson S, Gao WN, Man MQ, Kriehuber E, et al. Barrier recovery is impeded at neutral pH, independent of ionic effects: implications for extracellular lipid processing. *Arch Dermatol Res* 1998; 290: 215–222.
  26. Bouwstra JA, Gooris GS, Dubbelaar FE, Weerheim AM, Ponc M. pH, cholesterol sulfate, and fatty acids affect the stratum corneum lipid organization. *J Invest Dermatol* 1998; 3: 69–74.
  27. Friberg SE. Micelles, microemulsions, liquid crystals, and the structure of stratum corneum lipids. *J Soc Cosmet Chem* 1990; 41: 155–171.
  28. Osborne DW, Friberg SE. Role of stratum corneum lipids as moisture retaining agent. *J Dispers Sci Technol* 1987; 8: 173–179.
  29. Hatano Y, Man M-Q, Uchida Y, Crumrine D, Scharschmidt TC, Kim EG, et al. Maintenance of an acidic stratum corneum prevents emergence of murine atopic dermatitis. *J Invest Dermatol* 2009; 129: 1824–1835.
  30. Choi EH, Man MQ, Xu P, Xin S, Liu Z, Crurine DA, et al. Stratum corneum acidification is impaired in moderately aged human and murine skin. *J Invest Dermatol* 2007; 127: 2847–2856.
  31. Fluhr JW, Mao-Qiang M, Brown BE, Hachem JP, Moskowitz DG, Demerjian M, et al. Functional consequences of a neutral pH in neonatal rat stratum corneum. *J Invest Dermatol* 2004; 123: 140–151.
  32. Hachem JP, Roelandt T, Schurer N, Pu X, Fluhr J, Guidelo C, et al. Acute acidification of stratum corneum membrane domains using polyhydroxyl acids improves lipid processing and inhibits degradation of corneodesmosomes. *J Invest Dermatol* 2010; 130: 500–510.
  33. Brattsand M, Egelrud T. Purification, molecular cloning, and expression of a human stratum corneum trypsin-like serine protease with possible function in desquamation. *J Biol Chem* 1999; 274: 30033–30040.
  34. Ekholm E, Egelrud T. Expression of stratum corneum chymotryptic enzyme in relation to other markers of epidermal differentiation in a skin explant model. *Exp Dermatol* 2000; 9: 65–70.
  35. Ekholm E, Brattsand M, Egelrud T. Stratum corneum tryptic enzyme in normal epidermis: A missing link in the desquamation process? *J Invest Dermatol* 2000; 114: 56–63.
  36. Hachem JP, Houben E, Crumrine D, Man MQ, Schurer N, Roelandt T, et al. Serine protease signaling of epidermal permeability barrier homeostasis. *J Invest* 2006; 126: 2074–2086.
  37. Leyden JJ, Stewart R, Kligman AM. Updated in vivo methods for evaluating topical antimicrobial agents on human skin. *J Invest Dermatol* 72: 165–170, 1979.
  38. Korting HC, Hubner K, Greiner K, Hamm G, Braun-Falco O. Differences in the skin surface pH and bacterial microflora due to the long-term application of synthetic detergent preparations of pH 5.5 and pH 7.0. Results of a crossover trial in healthy volunteers. *Acta Derm Venereol* 1990; 70: 429–431.
  39. Schitteck B, Hipfel R, Sauer B, Bauer J, Kalbacher H, Stevanovic S, et al. Dermicidin: a novel human antibiotic peptide secreted by sweat glands. *Nat Immunol* 2001; 2: 1133–1137.
  40. Chikakane K, Takahashi H. Measurement of skin pH and its significance in cutaneous diseases. *Clin Dermatol* 1995; 13: 299–306.
  41. Weller R, Price RJ, Ormerod AD. Antimicrobial effect of acidified nitrite on dermatophyte fungi, *Candida* and bacterial skin pathogens. *J Appl Microbiol* 2001; 90: 648–652.
  42. Seidenari S, Giusti G. Objective assessment of the skin of children affected by atopic dermatitis: a Study of pH, capacitance and TEWL in eczematous and clinically uninvolved skin. *Acta Derm Venereol* 1995; 75: 429–433.
  43. Eberlein-König B, Schäfer T, Huss-Marp J, Darsow U, Möhrenschrager M, Herbert O, et al. Skin surface pH, stratum corneum hydration, trans-epidermal water loss and skin roughness related to atopic eczema and skin dryness in a population of primary school children. *Acta Derm Venereol* 2000; 80: 188–191.
  44. Sparavigna A, Setaro M, Gualandri V. Cutaneous pH in children affected by atopic dermatitis and in healthy children: a multicenter study. *Skin Res Technol* 1999; 5: 221–227.
  45. Krien PM, Kermici M. Evidence for the existence of a self-regulating enzymatic process within the human stratum corneum: an unexpected role for urocanic acid. *J Invest Dermatol* 2000; 115: 414–420.
  46. Tanaka M, Okada M, Zhen YX, Inamura N, Kitano T, Shirai S, et al. Decreased hydration state of the stratum corneum and reduced amino acid content of the skin in patients with seasonal allergic rhinitis. *Br J Dermatol* 1998; 139: 618–621.
  47. Parkkinen MU, Kiistala R, Kiistala U. Sweating response to moderate thermal stress in atopic dermatitis. *Br J Dermatol* 1992; 126: 346–350.
  48. Fartasch M, Diepgen TL. The barrier function in atopic dry skin: disturbance of membrane-coating granule exocytosis and formation of epidermal lipids? *Acta Derm Venereol Suppl* 1992; 176: 26–31.
  49. Chapman SJ, Walsh A. Membrane-coating granules are acidic organelles which possess proton pumps. *J Invest Dermatol* 1989; 93: 466–470.
  50. Pilgram GS, Vissers DC, van der Meulen H, Pavel S, Lavrijsen SP, Bouwstra JA, Koerten HK. Aberrant lipid organization in stratum corneum of patients with atopic dermatitis and lamellar ichthyosis. *J Invest Dermatol* 2001; 117: 710–717.
  51. Hansson L, Bäckman A, Ny A, Edlund M, Ekholm E, Ekstrand Hammarström B, et al. Epidermal over expression of stratum corneum chymotryptic enzyme in mice: a model for chronic itchy dermatitis. *J Invest Dermatol* 2002; 118: 444–449.

52. Steinhoff M, Neisius U, Ikoma A, Fartasch M, Heyer G, Skov PS, et al. Proteinase-activated receptor-2 mediates itch: a novel pathway for pruritus in human skin. *J Neurosci* 2003; 23: 6176–6180.
53. Whiting RC, Sackitey S, Calderone S, Morely K, Phillips JG. Model for the survival of *Staphylococcus aureus* in non growth environments. *Int J Food Microbiol* 1996; 31: 231–243.
54. Gianuzzi L, Contrera E, Zaritzky N. Modeling the aerobic growth and decline of *Staphylococcus aureus* as affected by pH and potassium sorbate concentration. *J Food Prot* 1999; 62: 356–362.
55. Kumaran D, Eswaramoorthy S, Furey W, Sax M, Swaminathan S. Structure of staphylococcal enterotoxin C2 at various pH levels. *Acta Crystallogr D Biol Crystallogr* 2001; 57: 1270–1275.
56. Mempel M, Schmidt T, Weidinger S, Schnopp C, Foster T, Ring J, Abeck D. Role of *Staphylococcus aureus* surface associated proteins in the attachment to cultured HaCaT keratinocytes in a new adhesion assay. *J Invest Dermatol* 1998; 111: 452–456.
57. Öhman H, Vahlquist A. The pH gradient over the stratum corneum differs in x-linked recessive and autosomal dominant ichthyosis: a clue to the molecular origin of the “acid skin mantle”? *J Invest Dermatol* 1998; 111: 674–677.
58. Odds FC. Morphogenesis in *Candida*, with special reference to *C. albicans*. In: *Candida and candidosis. A review and bibliography*. 2nd edn. London: Bailliere Tindall, 1988: 42–58.
59. Buffo J, Herman MA, Soll DR. A characterization of pH-regulated dimorphism in *Candida albicans*. *Mycopathologia* 1984; 85: 21–30.
60. Scherwitz C. Ultrastructure of human cutaneous candidosis. *J Invest Dermatol* 1982; 78: 200–205.
61. Runeman B, Faergemann J, Larkö O. Experimental *Candida albicans* lesions in healthy humans: dependence on skin pH. *Acta Derm Venereol* 2000; 80: 421–424.
62. Weller R, Ormerod AD, Hobson RP, Benjamin NJ. A randomized trial of acidified nitrite cream in the treatment of tinea pedis. *J Am Acad Dermatol* 1998; 38: 559–563.
63. Yosipovitch G, Tur E, Cohen O. Skin surface pH in intertriginous areas in NIDDM patients. Possible correlation to candidal intertrigo. *Diabetes Care* 1993; 16: 560–563.
64. Adam, R. Skin Care of the Diaper Area. *Pediatr Dermatol* 2008; 25: 427–433.
65. Berg RW, Milligan MC, Sarbaugh FC. Association of skin wetness and pH with diaper dermatitis. *Ped Dermatol* 1994; 11: 18–20.
66. Beguin AM, Malaquin-Pavan E, Guihaire C, Hallet-Lezy AM, Souchon S, Homann V, et al. Improving diaper design to address incontinence associated dermatitis. *BMC Geriatr* 2010; 10: 86.
67. Adam R, Schnetz B, Mathey P, Pericoi M, de Prost Y. Clinical demonstration of skin mildness and suitability for sensitive infant skin of a new baby wipe. *Pediatr Dermatol* 2009; 26: 506–513.
68. <http://www.rephreshbrilliant.com>
69. Seidenari S, Francomano M, Mantovani L. Baseline biophysical parameters in subjects with sensitive skin. *Contact Dermatitis* 1998; 38: 311–315.
70. Wilhelm KP, Maibach HI. Susceptibility to irritant dermatitis induced by sodium lauryl sulphate. *J Am Acad Dermatol* 1990; 23: 123–124.
71. Korting HC, Braun-Falco O. The effect of detergents on skin pH and its consequences. *Clin Dermatol* 1996; 14: 23–27.
72. Korting HC, Ponce-Pöschl E, Klövekorn W, Schmötzer G, Arens-Corell M, Braun-Falco O. The influence of the regular use of a soap or an acidic syndet bar on pre-acne. *Infection* 1995; 23: 89–93.
73. Yosipovitch G, Tur E, Morduchowicz, Boner G. Skin surface pH, moisture, and pruritus in haemodialysis patients. *Nephrol Dial Transplant* 1993; 8: 1129–1132.
74. Bencini PL, Montagnino G, Citterio A, Graziani G, Crosti C, Ponticelli C. Cutaneous abnormalities in uremic patients. *Nephro* 1985; 40: 316–321.
75. Rippke F, Schreiner V, Doering T, Maibach H. Stratum corneum pH in atopic dermatitis: impact on skin barrier function and colonization with *staphylococcus aureus*. *Am J Clin Dermatol* 2004; 5: 217–223.
76. Ananthapadmanabhan KP, Moore DJ, Subramanian K, Misra M, Meyer F. Cleansing without compromise: the impact of cleansers on the skin barrier and the technology of mild cleansing. *Dermatologic Therapy* 2004; 17: 16–25.
77. Ertel KD, Keswick BH, Bryant PB. A forearm controlled application technique for estimating the relative mildness of personal cleansing products. *J Soc Cosmet Chem* 1995; 46: 67–76.
78. Frosch PJ, Kligman AM. The soap chamber test. *J Am Acad Dermatol* 1979; 1: 35–41.
79. Sharko PT, Murahata RI, Leyden JL, Grove GL. Arm wash with instrumental evaluation – a sensitive technique for differentiating the irritation potential of personal washing products. *J Dermoclin Eval Soc* 1991; 2: 19–27.
80. Simion FA. Human in vivo methods for assessing the irritation potential of cleansing systems. In: MM Riger, LD Rhein, editors. *Surfactants in Cosmetics. Surfactant Science Series*, Vol. 68. New York, NY: Marcel Dekker, 1997: 519–532.
81. Simion FA, Rhein LD, Morrison BM Jr, Scala DD, Salko DM, Kligman AM, Grove GL. Self-perceived sensory response to soap and synthetic detergent bars correlate with clinical signs of irritation. *J Am Acad Dermatol* 1995; 32: 205–211.
82. Strube DA, Koontz SW, Murahata RI, Theiler R. The flex wash test: a method for evaluating the mildness of personal washing products. *J Soc Cosmet Chem* 1989; 40: 297–306.
83. Ananthapadmanabhan KP, Lips A, Vincent C, Meyer F, Caso S, Johnson A, et al. pH-induced alterations in stratum corneum properties. *Int J Cosmet Sci* 2003; 25: 103–112.
84. Baranda L, González-Amaro R, Torres-Alvarez B, Alvarez C, Ramírez V. Correlation between pH and irritant effect of cleansers marketed for dry skin. *Int J Dermatol* 2002; 41: 494–499.
85. Shi V, Tran K, Lio P. A comparison of physicochemical properties of a selection of modern moisturizers: hydrophilic index and pH. *J Drugs Dermatol* 2012; 11: 633–636.
86. Rawlings AV, Davies A, Carlomusto M, Pillai S, Zhang K, Kosturko R, et al. Effect of lactic acid isomers on keratinocyte ceramide synthesis, stratum corneum lipid levels and stratum corneum barrier function. *Arch Dermatol Res* 1996; 288: 383–390.
87. Imokawa G, Abe A, Jin K, Higaki Y, Kawashima M, Hidano A. Decreased levels of ceramides in stratum corneum of atopic dermatitis: an etiological factor in atopic dry skin? *J Invest Dermatol* 1991; 96: 523–526.
88. Yamamoto A, Serizawa S, Ito M, Sato Y. Stratum corneum lipid abnormalities in atopic dermatitis. *Arch Dermatol Res* 1991; 283: 219–223.
89. Sasai-Takedatsu M, Kojima T, Yamamoto A, Hattori K, Yoshijima S, Taniuchi S, et al. Reduction of *Staphylococcus aureus* in atopic skin lesions with acid electrolytic water: a new therapeutic strategy for atopic dermatitis. *Allergy* 1997; 52: 1012–1016.
90. Kubota K, Machida I, Tamura K, Take H, Kurabayashi H,



- Akiba T, Tamura J. Treatment of refractory cases of atopic dermatitis with acidic hot-spring bathing. *Acta Derm Venereol* 1997; 77: 452–454.
91. Fluhr, JW, Bankova L, Dikstein S. Skin surface pH, mechanism, measurement, importance. In: *Handbook of Non Invasive Methods and the Skin*, second edn. Ed. Serup J, Jemec G, Grove L, editors. Taylor & Francis Boca Raton 2006: p. 411–420.