

The pH of the Skin Surface and Its Impact on the Barrier Function

M.-H. Schmid-Wendtner^a H.C. Korting^b

Departments of Dermatology and Allergology, ^aRheinische Friedrich Wilhelm University, Bonn, and

^bLudwig Maximilians University, Munich, Germany

Key Words

Acid mantle · Barrier function · Skin cleansing · Skin pH · Synthetic detergents

Abstract

The 'acid mantle' of the stratum corneum seems to be important for both permeability barrier formation and cutaneous antimicrobial defense. However, the origin of the acidic pH, measurable on the skin surface, remains conjectural. Passive and active influencing factors have been proposed, e.g. eccrine and sebaceous secretions as well as proton pumps. In recent years, numerous investigations have been published focusing on the changes in the pH of the deeper layers of the stratum corneum, as well as on the influence of physiological and pathological factors. The pH of the skin follows a sharp gradient across the stratum corneum, which is suspected to be important in controlling enzymatic activities and skin renewal. The skin pH is affected by a great number of endogenous factors, e.g. skin moisture, sweat, sebum, anatomic site, genetic predisposition and age. In addition, exogenous factors like detergents, application of cosmetic products, occlusive dressings as well as topical antibiotics may influence the skin pH. Changes in the pH are reported to play a role in the pathogenesis of skin diseases like irritant contact dermatitis, atopic dermati-

tis, ichthyosis, acne vulgaris and *Candida albicans* infections. Therefore, the use of skin cleansing agents, especially synthetic detergents with a pH of about 5.5, may be of relevance in the prevention and treatment of those skin diseases.

Copyright © 2006 S. Karger AG, Basel

Introduction

The acidic nature of the skin surface was first determined by Heuss in 1892 [1]. Since then numerous studies have confirmed this observation. In 1928, Schade and Marchionini published their experiments on the skin pH using more exact biophysical methods. They introduced the 'gas chain bell electrode' adapted for the skin [2]. The acidic skin pH was interpreted as being due to impregnation of the horny layer with acidic constituents or eccrine sweat, because in intertriginous areas and in regions supplied by apocrine glands the reaction was less acidic or even neutral. Potentiometric measurements carried out by Blank [3] revealed skin pH values between 4.2 and 5.6. Since the beginning of the 1950s, the flat glass electrode was used more and more for skin pH measurements [4]. Schirren showed the equivalence between the plate glass electrode and the quinhydrone electrode measurement facing reproducible pH values around 5.2 for both meth-

Table 1. Factors influencing skin pH according to Rippke et al. [16], Yosipovitch and Maibach [18] and Jacobi et al. [25]

Endogenous factors
Age
Anatomic site
Genetic predisposition
Ethnic differences
Sebum
Skin moisture
Sweat
Exogenous factors
Detergents, cosmetic products, soaps
Occlusive dressings
Skin irritants
Topical antibacterials

ods [4]. On the basis of different experimental methods, Tronnier [5] suggested pH values closer to the alkaline range (pH 6.4–6.5), while Zlotogorski [6] even more recently reported pH values between 4.0 and 4.9. The overall evaluation of publications which are generally recognized today suggests a surface pH value of the human skin (forearm of a healthy adult white male) of 5.4–5.9 [7]. However, there are ‘physiological gaps in the acid barrier’ which deserve special attention. This refers in particular to the axillae, the genitoanal region and the interdigital area. Due to the alkaline environment, the normal bacterial flora in these areas is considered different.

Newer results concerning factors influencing the acid mantle of the skin as well as the role of skin pH in different skin diseases are the subject of this review.

Definition and pH Measurement

The pH is defined as the negative logarithm (base ten) of the concentration of free hydrogen ions in aqueous solution. The neutral point is 7 and the maximum values of the acidic and alkaline ranges are 0 and 14, respectively. The pH of the extractable water-soluble components of the skin can be measured. The skin pH and the buffering capacity of the skin surface are made up of the contributions from all the components of the stratum corneum as well as the secretions from sebaceous and sweat glands. Today, potentiometric methods, especially with flat glass electrodes, are preferred for skin pH measurements, since they are more accurate compared with colorimetric methods [8].

Barrier Function

The formation of the stratum corneum barrier involves several pH-dependent enzymes, especially with respect to its lipophilic components and its destruction by desquamation. These hydrolases include β -glucocerebrosidase, acid sphingomyelinase, acid lipases, phosphatases and phospholipases [9]. β -Glucocerebrosidase with an optimum pH of 5.6 is involved in the synthesis of the most important ceramides.

At the stratum granulosum/stratum compactum interface there is a release of acid hydrolases (contents of the lamellar bodies) into the extracellular space by differentiated keratinocytes. Together with those hydrolases, glucosylceramides and phospholipids play an important role in the lamellar arrangement of barrier lipids [10]. In vitro experiments showed that the formation of lamellar structures in mixtures of horny lipids requires an acidic milieu [11]. This observation was confirmed by in vivo studies in hairless mice which showed a significantly faster recovery of the barrier function after acetone insult or adhesive film-stripping when exposed to more acidic buffer solutions [10]. Leyvraz et al. investigated the epidermal barrier function in dependence on proteolytic enzymes [12]. They regenerated mice lacking the membrane-anchored channel-activating serine protease (CAP 1) in skin, and these mice died within 60 h after birth. Besides a lower body weight they exhibited severe malformation of the stratum corneum, which was accompanied by an impaired skin barrier function. As evidenced by dehydration and skin permeability assay and transepidermal water loss measurements, this resulted in a rapid and fatal dehydration [12].

Various investigations revealed that low pH values in the extracellular space play an important role in the regulation of enzyme activity, especially in keratinization and barrier regeneration [13–15]. In this respect, acidic buffer substances applied topically may contribute to the maintenance of the normal barrier function of the skin.

Factors Affecting Skin pH

The pH of the skin surface can be influenced by a number of endogenous (physiological) and exogenous factors [16–18]. The most important of these (table 1) are discussed in more detail.

Age has long been known to influence the skin pH. Previous studies indicated that the skin undergoes a process of adaptation and maturation postnatally. In recent

years, experimental studies especially focused on infants, small children and the elderly. Visscher et al. [19] reported relatively neutral surface pH values in neonates with a decrease to about 5.5 over the first 4 postnatal days. These data were confirmed by Yosipovitch et al. [20] who measured elevated initial mean pH values of 7.08 (SD 0.17) with significant decreases of pH by day 2. Hoeger and Enzmann [21] assessed skin function parameters (skin surface pH, corneal layer hydration, epidermal desquamation, and surface roughness) prospectively in a cohort of 202 healthy term neonates. They found a decrease in surface pH by 0.3–1.1 units, while desquamation increased significantly during the observation period (12 weeks). In addition, stratum corneum hydration increased significantly, paralleled by a decrease in skin roughness [21]. Data concerning the skin pH of schoolchildren are inhomogeneous. There might be a reduced resistance to alkaline noxae despite an unremarkable basal pH. In contrast, both an increased skin surface pH and a reduced buffer capacity have been documented for skin of the elderly [22]. In the 18–60-year age group, the skin surface pH remains constant in most anatomical sites [22].

Comparing data obtained from different anatomical sites may often be inappropriate because skin surface composition is not necessarily uniform and hygiene habits may be totally different depending on the body area. However, it is generally accepted that skin surface pH is relatively similar at different body sites except in areas with higher moisture: the intertriginous areas (the axilla, inguinal and submammary folds and finger webs) have a slightly higher pH than other body areas [7, 23].

Skin pH has occasionally been reported to vary with ethnic and genetic background. However, racial differences in skin have been minimally investigated by objective methods. It has been demonstrated that people with black skin have a lower skin surface pH than people with white skin [24]. The assumption that brown-skinned individuals from India might have a slightly different skin surface pH compared with Europeans has not yet been proven by exact statistical methods [17]. Studies concerning gender-related differences in skin physiology provided conflicting results. A newer investigation involving 6 female and 6 male volunteers revealed a higher pH for women (5.6 ± 0.4) than for men (4.3 ± 0.4 , $p < 0.05$) [25].

The sebum of the skin has a moderate effect on the pH to be measured. The effect depends on the amount of sebum. For example, a high sebum content on the forehead in a patient with acne vulgaris may artificially affect the pH measurement. Skin pH also depends on skin mois-

ture: areas with higher moisture such as the axilla and inguinal regions have a higher pH.

In historic reports, differences in sweat pH according to the sweating intensity are reported [26]. Until today there is no confirmation that thermal sweat, sweat from covered body areas and sweat from eccrine glands really is more acidic than sweat from effort, air-exposed skin areas and apocrine glands.

There are numerous external factors affecting the measured skin surface pH. One of the most important ones is skin cleansing. A pH rise for few hours at least is observed after cleansing the skin with alkaline soaps (pH 10.5–11.0). The use of synthetic detergents formulated at the same pH as skin and even tap water also leads to a rise of the skin surface pH, however to a lesser extent and for a shorter period of time [27, 28]. It is assumed that such temporary skin pH changes are limited to the uppermost layers of the stratum corneum, which was examined using the tape-stripping method [17]. About 15 years ago, Korting et al. [29, 30] were one of the first groups who examined the effect of different skin cleansing methods in healthy volunteers focusing on the bacterial flora and the skin surface pH. In a clinical crossover study one half of the healthy volunteers washed their foreheads and forearms twice daily for 1 min with soap, the other volunteers washed with a syndet with an acidic pH. After 4 weeks, the respective other skin cleansing agent was used for the next 4 weeks. At the beginning of the study and at the end of every week, skin pH and density of the bacterial flora were determined [29]. When soap was used first, the pH increased. After change-over to the syndet the pH dropped. When the syndet was applied first, the pH dropped slightly, but after changing to soap it increased to values above the initial values [29]. Statistical evaluations showed that the pH values were about 0.3 units lower in the case of long-term application of the acidic syndet. However, after individual washing, both the soap and the syndet led to a short-term increase in pH, which regressed within a few hours [26]. After repeated soap washing, the count of propionibacteria increased strongly and dropped after repeated syndet application. Moreover, for the forehead a correlation could be established between germ species and the skin surface pH [29]. Various topical products, for example deodorants, have been reported to affect the skin surface pH, at least for a few hours [31, 32]. The effects of occlusive dressings on the skin surface pH have been known for several decades. Hartmann [33] demonstrated a significant increase in the skin surface pH in healthy volunteers after 3 days of occlusion with a return to the baseline pH

value only 1 day after removal of the occlusive dressing. Elsner and Maibach [34] reported a significant decrease in the skin pH after drying of primary occluded skin of the vulva, associated with a significant reduction of the transepidermal water loss. This was explained by an increased ion permeability of the stratum corneum because of occlusion, resulting in neutralization of the acidic skin surface [34]. In addition, a delay in restoration of the barrier function after occlusive acetone treatment was described associated with a rise in the skin surface pH [35].

Skin pH in Diseases

Both lipid organization and lipid metabolism in the stratum corneum require an acidic pH. Alterations in those processes contribute to the disturbance of skin barrier function observed in different skin diseases. Today we know that a disturbed stratum corneum skin pH has important clinical implications in several clinical situations, for example in psoriasis and atopic dermatitis [36] as well as in more rare dermatoses like the Netherton syndrome [37]. Frequent skin diseases with disturbed skin pH are discussed in more detail below.

Atopic Dermatitis

Several pathogenetic and therapeutically relevant factors in atopic dermatitis can be affected by changes in skin surface pH. Besides genetic and immunologic factors, skin dryness caused by a disturbance of skin barrier function and microbial colonization may play an important role. About 50 years ago there was initial evidence for an altered skin surface pH in patients with atopic dermatitis. Epprecht [38], for example, described elevated pH values of the skin surface, with slower alkaline neutralization, in patients with acute disseminated atopic dermatitis. Schnyder et al. [39] determined a reduced alkaline resistance in 83% of patients with atopic dermatitis compared with 17% of those with healthy skin. These early data have been confirmed in recent years by numerous systematic studies, using planar glass electrodes for pH measurement [40, 41].

Seidenari and Giusti [41] showed that the pH values measured in eczematous and in clinically uninvolved skin areas were significantly higher in patients with atopic dermatitis than those in healthy children. In addition, the pH values in eczematous skin areas were significantly higher than in unaffected areas, which in turn were significantly higher in patients with clinical findings than

in patients in a symptom-free interval [41]. These findings were corroborated by Sparavigna et al. [42], who compared the cutaneous pH of 143 children with atopic dermatitis and 141 healthy children, aged 3 months to 11 years. They reported a progressive decrease in pH values from lesional to perilesional to unaffected skin sites, and to corresponding skin sites of healthy children [42]. Eberlein-König et al. [40] examined 377 primary schoolchildren for skin surface pH, stratum corneum hydration, transepidermal water loss and skin roughness. Only the skin pH differed significantly, with higher mean values in the 45 children with atopic dermatitis compared with the healthy children. Furthermore, an elevated pH could be correlated with the severity of dryness and itching [40].

Whereas the significance of free fatty acids for the alterations of pH in atopic skin remains inconclusive [43], recent findings suggest that urocanic acid may play a key role in maintenance of the acidic pH of the stratum corneum [44]. According to the authors, urocanic acid is produced from histidine by a self-regulated control cycle, involving the pH-dependent activity of the enzyme histidase for the maintenance of moisture content and pH of the stratum corneum. Other studies confirmed a significant reduction in urocanic acid and suggested a diminished histidase activity in atopic skin [45].

Irritant Contact Dermatitis

The pH of the skin surface can also be used to predict the risk of individuals without atopic background to develop irritant contact dermatitis. Wilhelm and Maibach [46] compared several skin parameters in 10 healthy volunteers before and after inducing skin irritation with 1% sodium lauryl sulphate. The authors reported significant correlations for the surface pH, the pH of the deeper stratum corneum layers exposed after five tape stripings, but not for the transepidermal water loss, sebum content or renewal rate of the horny layer [46]. These findings were confirmed by the group of Francomano [47] who observed higher pH values of the cheeks in patients with sensitive skin objectively prone to skin irritations, than in healthy volunteers. Disturbances of the skin pH play a pathogenetic role in infants, too. Ammonia-induced alkalization activates stool enzymes such as lipase and trypsin, leading to irritation and disruption of the skin barrier, where the skin has been reported in some cases to already have an elevated baseline pH of 6.6 [48].

Ichthyosis

Öhman and Valquist [49] compared the pH gradients of the horny layer of patients with X-chromosomal recessive ichthyosis and autosomal dominant ichthyosis vulgaris with those of healthy volunteers. The pH on the skin surface in patients with ichthyosis vulgaris was significantly higher (5.3, SD 0.7) than in patients with X-chromosomal recessive ichthyosis (4.6, SD 0.4), and was higher than the pH of healthy skin (4.5, SD 0.2). A neutral pH was reached in patients with ichthyosis vulgaris after half the horny layer had been removed using the tape stripping method. In contrast, in patients with X-chromosomal recessive ichthyosis, a plateau pH value of 6.2–6.6 was never exceeded even in the lower stratum corneum. It was concluded that in patients with ichthyosis vulgaris there is a congenital deficiency of acid degradation products of filaggrin, whereas in patients with X-chromosomal recessive ichthyosis there is an accumulation of cholesterol sulphate in the lower horny layer induced by a lack of steroid sulphatase, leading to a flattening of the pH gradient [49]. As a result, pH-dependent enzymes involved in desquamation of the skin may be disturbed. Reduced filaggrin formation levels have been confirmed in patients with ichthyosis vulgaris, lamellar ichthyosis and congenital ichthyosiform erythrodermia [50]. The steep amino acid gradient, with high values in the deep horny layer of healthy skin, is much flatter in ichthyosis vulgaris. This might explain the advantage of special acidic topical preparations containing lactic acid for keratolysis in patients with ichthyosis vulgaris.

Fungal Infections in Special Patient Populations

Yosipovitch et al. [51] compared skin surface pH and moisture in intertriginous areas in 50 non-insulin-dependent diabetic patients and healthy volunteers. They measured significantly higher pH values in the intertriginous skin areas of these patients (but not on the forearms) compared with the healthy control group. This was interpreted as a risk factor for the development of candida mycoses [51]. In another investigation, a significant increase in the skin pH in patients on dialysis was reported [52]. The changes observed, relating to the tendency to develop mycotic skin infections, were considered to have clinical relevance [52]. In an experimental setting, *Candida albicans* cell suspension, buffered either at pH 6.0 or 4.5, was applied to healthy skin under occlusion. After 24 h, there were significantly less mycotic skin lesions at the acidic site, which confirms the relevance of an intact acidic milieu for the patient's defense capability against fungal infections [53].

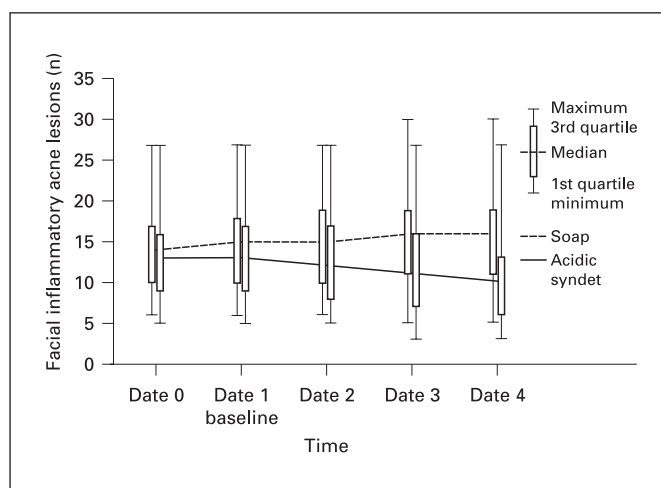


Fig. 1. Number of facial inflammatory acne lesions in a comparative trial with 3-month application of either soap or acidic syndet (from Korting et al. [57], with permission).

Skin pH and Skin Cleansing

Even rinsing the skin with water alone immediately produces a transient increase in the skin pH [54]. Washing the hands with conventional soap causes the pH on the palms to increase by an average of 3 units. Even 90 min after washing with soap the pH of the hands was not completely normalized [55]. However, soaps were used for over thousands of years for regular skin cleansing, while synthetic cleansers of the liquid or the bar type have started to become an alternative just about 30 years ago. Keining [56] was the first to describe the advantages of the so-called syndets. He mentioned a decreased irritancy potential, lack of sensitization and capability to maintain or even restore the acid mantle of the skin [56]. Today, the value of acidic syndets in the therapy of various skin diseases is widely accepted on clinical grounds. For quite a while, a close relationship has been postulated between skin surface pH and its microbial flora (transient and resident). Bacteria can grow over a wide pH range, but no microorganisms will grow equally at all pH values. Most grow better at a pH around neutrality. Acidic pH could be bacteriostatic for some strains. Korting et al. [57], for example, investigated the influence of the regular use of a soap or an acidic syndet bar on acne-prone patients. In a randomized, open-labeled, comparative trial the 3-month application of either skin cleansing preparation to facial skin for 1 min each in the morning and in the evening was compared in 120 adolescents and young adults with inflammatory acne I or II accord-

ing to the Plewig and Kligman classification [57]. The number of inflammatory acne lesions, i.e. papulopustules, differed in the two trial groups from 4 weeks of application onward. In the group using soap the mean number of inflammatory lesions increased from 14.6 (SD 5.3) to 15.3 (SD 6.0), while it decreased in the other group from 13.4 (SD 5.2) to 10.4 (SD 5.8; $p < 0.0001$; fig. 1). Acidic syndet bars, in addition, showed a better tolerability. The authors concluded that acidic cleansers should be preferred for skin care in acne-prone adolescents and young adults.

Just recently, the impact of acidic hand cleansers for prevention of rhinovirus infections has been investigated, since direct hand-to-hand contact is an important mechanism of transmission of rhinovirus infection [58]. Hands of volunteers were contaminated with rhinovirus at defined times (15 min, 1 h and 3 h) after use of acidic hand cleansers (containing salicylic acid and/or pyroglutamic acid). Then volunteers attempted to inoculate the nasal mucosa with one hand while quantitative viral cultures were done on the other hand. Virus was recovered from the hands in 90% of the volunteers in the control group compared with 15% in the groups who used acidic hand cleansers after 15 min. Rhinovirus infection occurred in 32% of patients in the control group and in only 7% of patients in the treatment groups. The authors suggested the feasibility of the prevention of rhinovirus transmission by acidic hand treatments that are virucidal on contact and have activity that persists after application [58].

References

- 1 Heuss E: Die Reaktion des Schweißes beim gesunden Menschen. *Monatschr Prakt Dermatol* 1892;14:341–343.
- 2 Schade H, Marchionini A: Der Säuremantel der Haut (nach Gaskettenmessung). *Klin Wochenschr* 1928;7:12–14.
- 3 Blank HI: Measurement of pH of the skin surface. *J Invest Dermatol* 1939;2:67–79.
- 4 Schirren CG: Does the glass-electrode determine the same pH values on the skin surface as a quinhydrone electrode? *J Invest Dermatol* 1955;24:485–488.
- 5 Tronnier A: Seifen und Syndets in der Hautpflege und Therapie. *Ärztl Kosmetol* 1985;15:19–30.
- 6 Zlotogorski A: Distribution of skin surface pH on the forehead and cheek of adults. *Arch Dermatol Res* 1987;398–401.
- 7 Braun-Falco O, Korting HC: Der normale pH-Wert der Haut. *Hautarzt* 1986;37:126–129.
- 8 Dikstein S, Zlotogorski A: Measurements of skin pH. *Acta Derm Venereol (Stockh)* 1994;(suppl 185):18–20.
- 9 Redoules D, Tarroux R, Perie J: Epidermal enzymes: their role in homeostasis and their relationship with dermatoses. *Skin Pharmacol Appl Skin Physiol* 1998;11:183–192.
- 10 Mauro T, Holleran WM, Grayson S, et al: Barrier recovery is impeded at neutral pH, independent of ionic effects: implications for extracellular lipid processing. *Arch Dermatol Res* 1998;290:215–222.
- 11 Bouwstra JA, Gooris GS, Dubhelaar FER, et al: pH, cholesterol sulfate, and fatty acids affect the stratum corneum lipid organization. *J Invest Dermatol* 1998;3:69–74.
- 12 Leyvraz C, Charles RP, Rubera I, et al: The epidermal barrier function is dependent on the serine protease CAP 1/Prss8. *J Cell Biol* 2005;170:487–496.
- 13 Vahlquist A: Variations in skin pH during normal and pathological keratinisation. *Retinoids* 1999;139:855–857.
- 14 Fluhr JW, Kao J, Jain M, et al: Generation of free fatty acids from phospholipids regulates stratum corneum acidification and integrity. *J Invest Dermatol* 2001;117:44–51.
- 15 Elias P: Stratum corneum architecture, metabolic activity and interactivity with subjacent barrier function. *Br J Dermatol* 1996;5:191–201.
- 16 Rippke F, Schreiner V, Schwanitz HJ: The acidic milieu of the horny layer. New findings on the physiology and pathophysiology of skin pH. *Am J Clin Dermatol* 2003;3:261–272.
- 17 Parra JL, Paye M: EEMCO guidance for the in vivo assessment of skin surface pH. *Skin Pharmacol Appl Skin Physiol* 2003;16:188–202.
- 18 Yosipovitch G, Maibach HI: Skin surface pH: a protective acid mantle. *Cosmet Toiletries* 1996;111:101–102.

Conclusion

A central role for the acidic milieu as a regulating factor in stratum corneum homeostasis was postulated. This has relevance for the integrity of the barrier function, from normal maturation of the stratum corneum lipids through to desquamation. An elevated skin pH may also influence the activity of the enzymatic process of lipid metabolism in the stratum corneum, which might contribute to a dramatic skin barrier anomaly observed, for example, in atopic dermatitis. In addition, changes in skin pH become clinically significant because of the creation of a favorable environment for the growth of bacteria, especially *Staphylococcus aureus*. Because of their bacteria-regulating properties and favorable tolerability profile, syndets with an acidic pH are now preferred for skin cleansing in patients with seborrheic-type diseases (acne vulgaris, rosacea), atopic skin diathesis, irritant contact dermatitis and ichthyosis. Moreover, special populations such as diabetic patients or patients on dialysis may profit from washing with acidic syndets because the occurrence of fungal infections may be reduced. There is good reason to believe that acidic syndets are of value even for persons with healthy skin, for example to prevent rhinovirus transmission and infection.

- 19 Visscher MO, Chatterjee R, Munson KA, et al: Changes in diapered and non-diapered infant skin over the first month of life. *Pediatr Dermatol* 2000;17:45–51.
- 20 Yosipovitch G, Maayan-Metzger A, Merlob P, et al: Skin barrier properties in different body areas in neonates. *Pediatrics* 2000;106:105–108.
- 21 Hoeger PH, Enzmann CC: Skin physiology of the neonate and young infant: a prospective study of functional skin parameters during early infancy. *Pediatr Dermatol* 2002;19:256–262.
- 22 Zlotogorski A: Distribution of skin surface pH on the forehead and cheek of adults. *Arch Dermatol Res* 1987;279:398–401.
- 23 Dikstein S, Zlotogorski A: Skin surface hydrogen ion concentration (pH); in Leveque JL (ed): *Cutaneous Investigation in Health and Disease*. New York, Marcel Dekker, 1989, pp 59–77.
- 24 Naissan O, Maibach W, Maibach HI: Racial (ethnic) differences in skin properties. *Am J Clin Dermatol* 2003;4:843–860.
- 25 Jacobi U, Gautier J, Sterry W, et al: Gender-related differences in the physiology of the stratum corneum. *Dermatology* 2005;211:312–317.
- 26 Talbert GA: Effect of work and heat on the hydrogen-ion concentration of the sweat. *Am J Physiol* 1919;50:433.
- 27 Korting HC, Braun-Falco O: The effects of detergents on skin pH and its consequences. *Clin Dermatol* 1995;14:23–27.
- 28 Barel AO, Lambrecht R, Clarys P, et al: A comparative study of the effects on the skin of a classical bar soap and a syndet cleansing bar in normal use conditions and in the soap chamber test. *Skin Res Technol* 2001;7:98–104.
- 29 Korting HC, Kober M, Müller M, et al: Influence of repeated washings with soap and synthetic detergents on pH and resident flora of the skin of forehead and forearm. Results of a cross-over trial in health probationers. *Acta Derm Venereol (Stockh)* 1987;67:41–47.
- 30 Korting HC, Greiner K, Hübner K, et al: Changes in the skin pH and resident flora by washing with synthetic detergent preparations at pH 5.5 and 8.5. *J Soc Cosmet Chem* 1991;42:147–158.
- 31 Stenzaly-Achttert S, Schölermann A, Schreiber J, et al: Axillary pH and influence of deodorants. *Skin Res Technol* 2000;6:87–91.
- 32 Wickett RR, Trobaugh CM: Personal care products: effect on skin surface pH. *Cosmet Toilet* 1990;105:41–46.
- 33 Hartmann AA: Effect of occlusion on resident flora, skin-moisture and skin-pH. *Arch Dermatol Res* 1983;275:251–254.
- 34 Elsner P, Maibach HI: The effect of prolonged drying on transepidermal water loss, capacitance and pH of human vulvar and forearm skin. *Acta Derm Venereol (Stockh)* 1990;70:105–109.
- 35 Schreiner V, Maerker U, Hoppe U: Dependence of barrier repair in human skin on intracellular and extracellular pH. *J Invest Dermatol* 1996;106:917.
- 36 Farwanah H, Raith K, Neubert RH, et al: Ceramide profiles of the uninvolved skin in atopic dermatitis and psoriasis are comparable to those of healthy skin. *Arch Dermatol Res* 2005;296:514–512.
- 37 Chao SC, Richard G, Lee JY: Netherton syndrome: report of two Taiwanese siblings with staphylococcal scaled skin syndrome and mutation of SPINK5. *Br J Dermatol* 2005;152:159–165.
- 38 Epprecht R: Elektrometrische Messungen mit besonderer Berücksichtigung der Säureneutralisation. *Dermatologica* 1955;111:204–223.
- 39 Schnyder UW, Gloor M, Taugner M: Über die sozialmedizinische Bedeutung von Alkaliresistenz, Alkalineutralisation und Hautoberflächenlipidmenge bei Neurodermitis atopica und Ichthyosis vulgaris. *Berufsdermatosen* 1977;25:101–140.
- 40 Eberlein-König B, Schäfer T, Huss-Marp J, et al: Skin surface pH, stratum corneum hydration, trans-epidermal water loss and skin roughness related to atopic eczema and skin dryness in a population of primary school children. *Acta Derm Venereol (Stockh)* 2000;80:188–191.
- 41 Seidenari S, Giusti G: Objective assessment of the skin of children affected by atopic dermatitis and in healthy children: a multicenter study. *Acta Derm Venereol (Stockh)* 1995;75:429–433.
- 42 Sparavigna A, Setaro M, Gualandri V: Cutaneous pH in children affected by atopic dermatitis and in healthy children: a multicenter study. *Skin Res Technol* 1999;5:221–227.
- 43 Hara J, Higuchi K, Okamoto R, et al: High-expression of sphingomyelin deacylase is an important determinant of ceramide deficiency leading to barrier disruption in atopic dermatitis. *J Invest Dermatol* 2000;115:406–413.
- 44 Krien PM, Kermici M: Evidence for the existence of a self-regulation enzyme process within the human stratum corneum: an unexpected role for urocanic acid. *J Invest Dermatol* 2000;115:414–420.
- 45 Abeck D, Lauer W, Schmidt T, et al: Age-dependent differences of histidine urocanic acid and urocanic isomers concentration in atopic eczema (abstract). *J Invest Dermatol* 1996;107:468.
- 46 Wilhelm KP, Maibach HI: Susceptibility to irritant dermatitis induced by sodium lauryl sulphate. *J Am Acad Dermatol* 1990;23:122–124.
- 47 Francomano MA, Mantovani L, Pepe P, et al: Baseline biophysical parameters in subjects with sensitive skin (abstract). *Skin Res Technol* 1996;2:225.
- 48 Gfatter R, Hackl P, Braun F: Effects of soap and detergents on skin surface pH, stratum corneum hydration and fat content in infants. *Dermatology* 1997;195:258–262.
- 49 Öhmann H, Vahlquist A: The pH gradient over the stratum corneum differs in X-linked recessive and autosomal dominant ichthyosis: a clue to the molecular origin of the acid skin mantle? *J Invest Dermatol* 1998;111:674–677.
- 50 Menon GK, Ghadially R, Williams ML, et al: Lamellar bodies as delivery systems of hydrolytic enzymes: implications for normal and abnormal desquamation. *Br J Dermatol* 1992;126:337–345.
- 51 Yosipovitch G, Tur E, Cohen O, et al: Skin surface pH in intertriginous areas in non-insulin-dependent diabetes mellitus patients. *Diabetes Care* 1993;16:560–563.
- 52 Yosipovitch G, Tur E, Morduchowitz G, et al: Skin surface pH, moisture, and pruritus in haemodialysis patients. *Nephrol Dial Transplant* 1993;8:1129–1132.
- 53 Runeman B, Faergemann J, Larkö O: Experimental *Candida albicans* lesions in healthy humans: dependence on skin pH. *Acta Derm Venereol* 2000;80:421–424.
- 54 Gfatter R, Hackl P, Braun F: Effects of soap and detergents on skin surface pH, stratum corneum hydration and fat content in infants. *Dermatology* 1997;195:258–262.
- 55 Mücke H, Mohr KT, Rümmler A, et al: Untersuchungen über den Haut-pH-Wert der Hand nach Anwendung von Seife, Reinigungs- und Händedesinfektionsmitteln. *Pharmazie* 1993;48:468–469.
- 56 Keining E: Zur Frage der Reinigung gesunder und kranker Haut. *Dermatol Wochenschr* 1959;46:1245–1251.
- 57 Korting HC, Ponce-Pöschl E, Klöveborn W, et al: The influence of the regular use of a soap or an acidic syndet bar on pre-acne. *Infection* 1995;23:89–93.
- 58 Turner RB, Biedermann KA, Morgan JM, et al: Efficacy of organic acids in hand cleansers for prevention of rhinovirus infections. *Antimicrob Agents Chemother* 2004;46:2595–2598.